

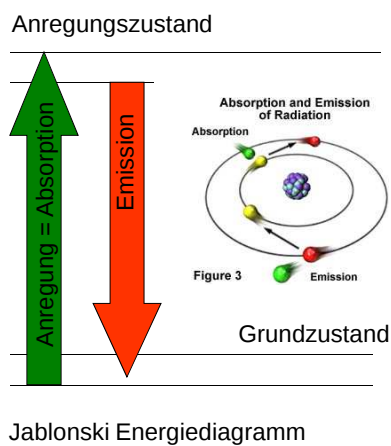
Fraunhofer IZI

Experimental Imaging - Alexander Kranz, MD

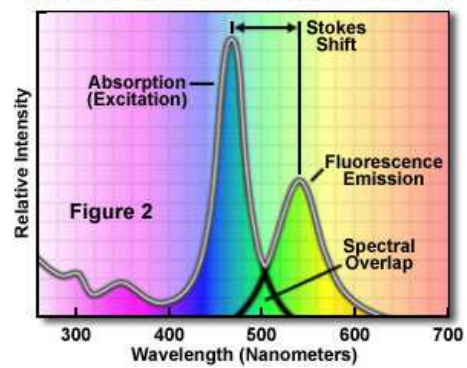


Fraunhofer
IZI

Basic Principle Fluorescence Imaging



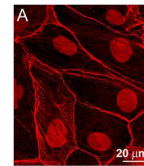
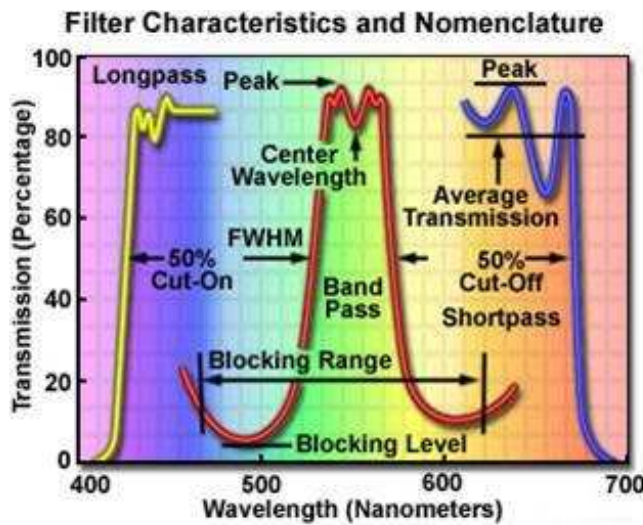
Excitation and Emission Spectral Profiles



Farbstoff	Exzitation	Emission
DAPI	359	461
FITC	495	519
Alexa 488	499	520

Fraunhofer
IZI

(Emission) filters

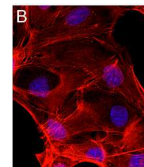


Excitation

543nm

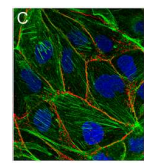
Beamsplitter:
HFP 488/543

Emission:
560nm LP



Emission:

560-615 nm BP
650 nm LP



Emission:

Spectral 11nm
552-723nm

North et al., 2006

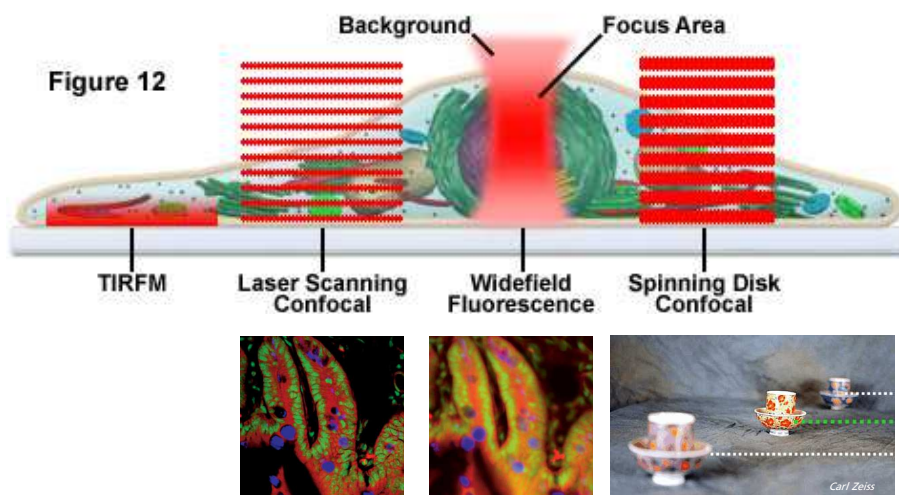
Ex/Em Filter must be selected to match your fluorochrome combination

Fraunhofer
IZI

Overview Fluorescence Imaging

Fluorescence Imaging Modes in Live-Cell Microscopy

Figure 12

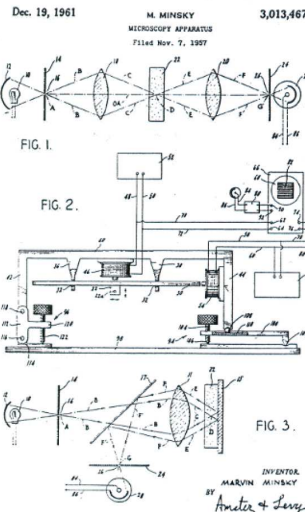
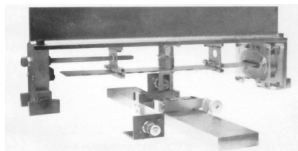


Fraunhofer
IZI



Marvin Minsky

1955



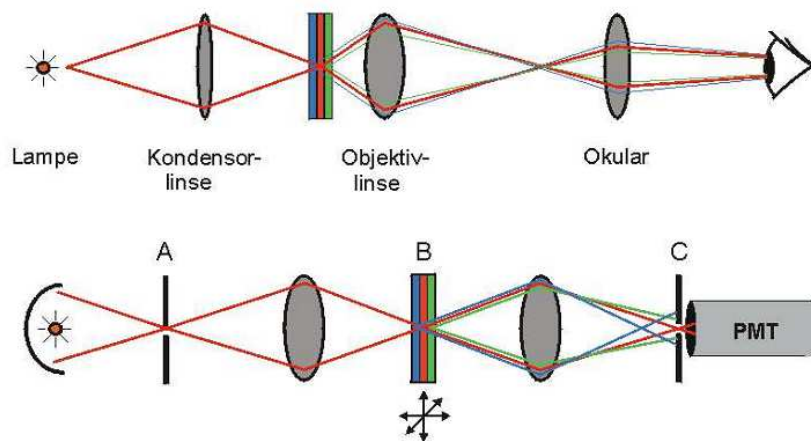
1988



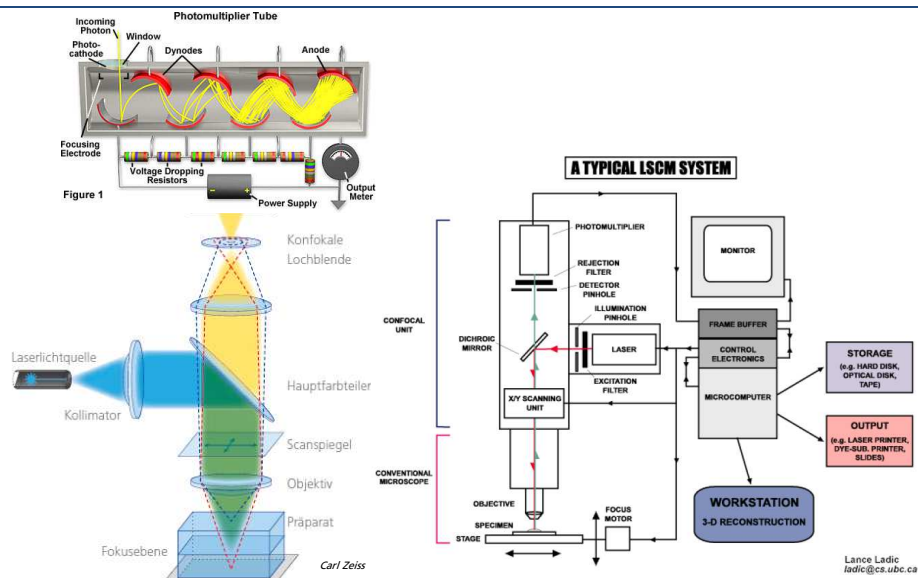
1992



2000

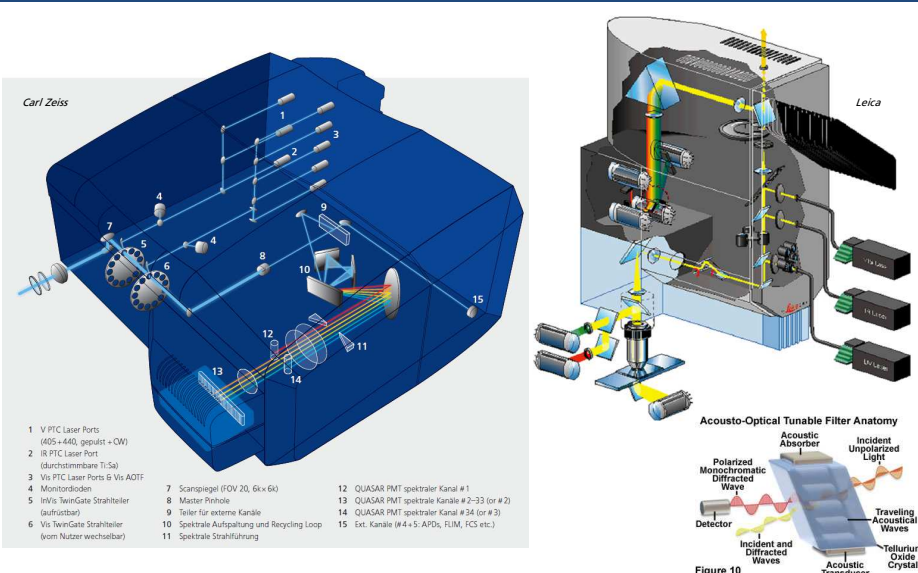


Composition



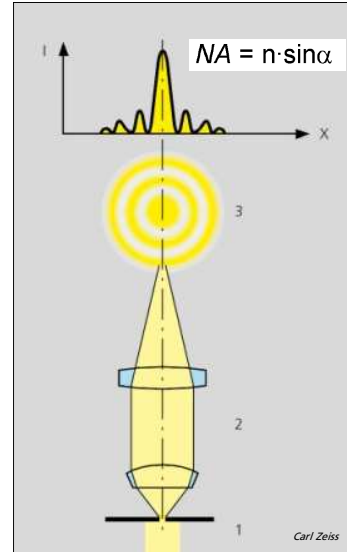
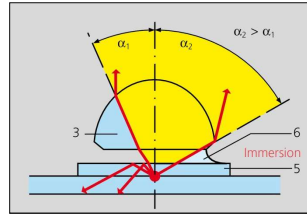
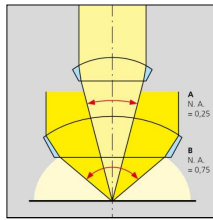
Fraunhofer
IZI

Optical path



Fraunhofer
IZI

Numeric aperture and resolution

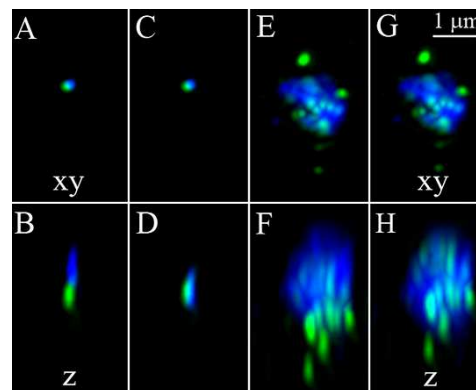
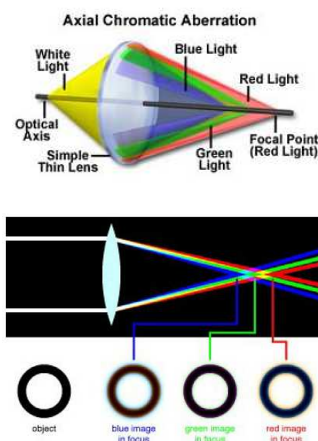


Öffnungswinkel des Objektivs
Brechungsindex des Immersionsmediums (n)

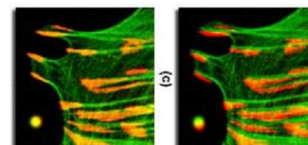
Definiert das Auflösungsvermögen
Kann in Luft NIE größer als 1 werden

Ein einzelner Punkt wird als Airy Scheibe abgebildet

Chromatic aberration

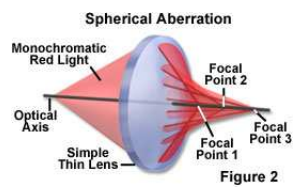


North et al., 2006

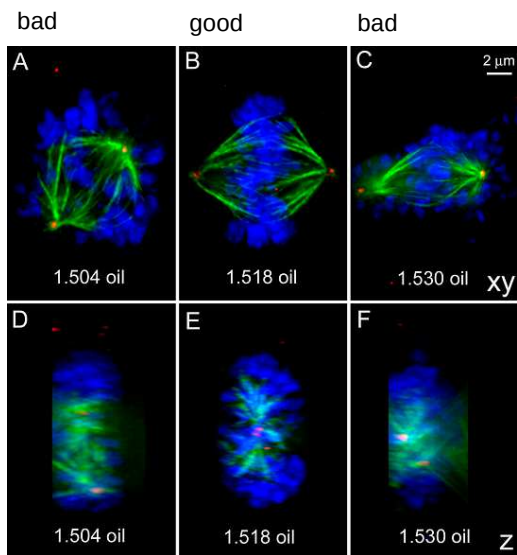


Correction with color beat

Spherical aberration



Refractive index mismatch
=wrong immersion medium
gives rise to geometrical
aberrations

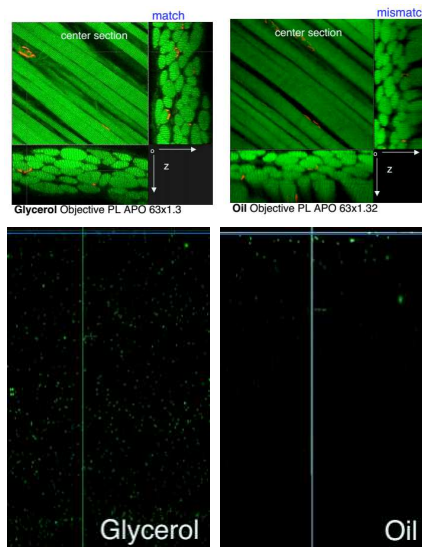
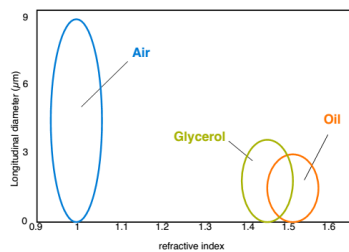


North et al., 2006

Fraunhofer
IZI

Mounting medium

Compound	Refractive Index	
Ethylenglycol	1.109	
Methanol	1.329	
Aqua	1.330	
Aceton	1.359	
Ethanol	1.361	
n-Hexan	1.375	
Isopropanol	1.378	
Glycerol in PBS pH7,4	1.385	40%
1-Propanol	1.385	
Mowiol	1.389	
Glycerol in PBS pH7,4	1.394	50%
Isobutanol	1.396	
Ethylenglycolmonomethylether	1.402	
Glycerol in PBS pH7,4	1.405	60%
Diethylglycoldiethylether	1.412	
Glycerol in PBS pH7,4	1.417	70%
Dioxan	1.422	
Glycerol in PBS pH7,4	1.425	80%
Glycerol in PBS pH7,4	1.435	90%
chloroform	1.448	



Fraunhofer
IZI

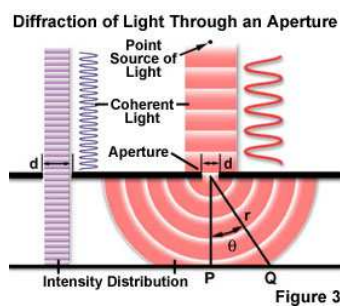
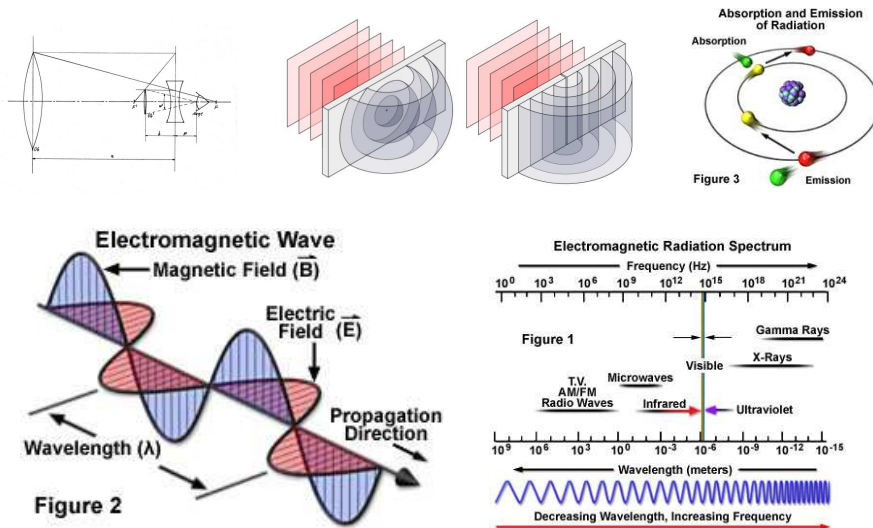


Figure 3

$$1AE = \frac{1,22\bar{\lambda}}{NA} = \text{Numerische Apertur}$$

$$\bar{\lambda} = \sqrt{2} \frac{\lambda_{em} \lambda_{exc}}{\sqrt{\lambda_{em}^2 + \lambda_{exc}^2}}$$

Airy Disk Patterns and PSFs from Diffraction

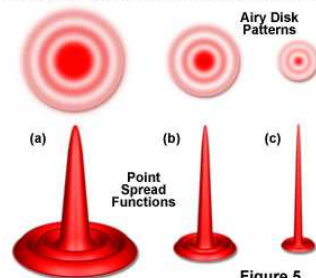
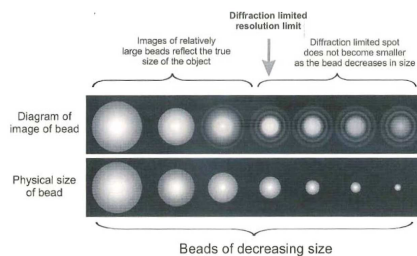
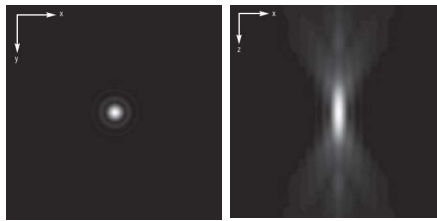


Figure 5

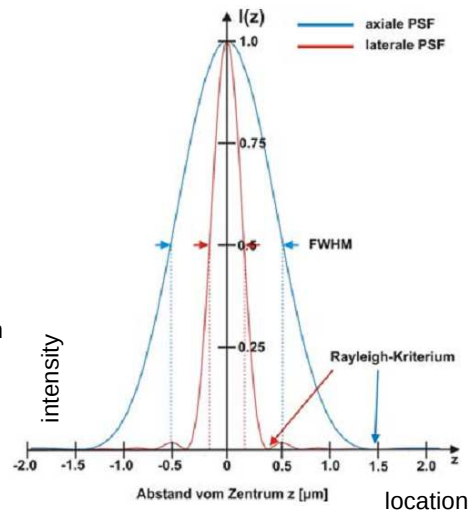


Point spread function



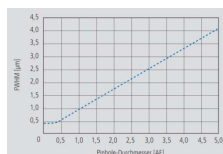
Laterale Projektion Axiale Projektion

- Point Spread Function (PSF)
- Wenn Intensität auf die Hälfte des Zentralen Maximum abgesunken dann Halbwertsfläche
- Setzt sich aus axialen und lateralen Anteil zusammen: Halbwertsbreite =
- Full Width at Half Maximum (FWHM)

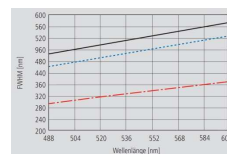


FWHM characteristics

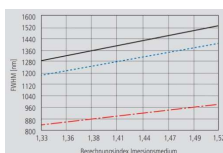
FWHM ist abhängig von:



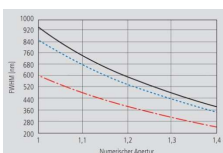
Blendengröße =
Pinhole
Durchmesser



Wellenlänge des
verwendeten
Lichts



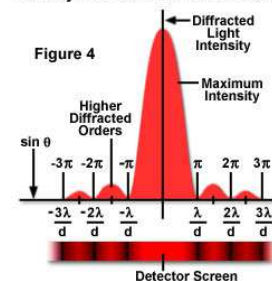
Brechungsindex
Imersionsmedium



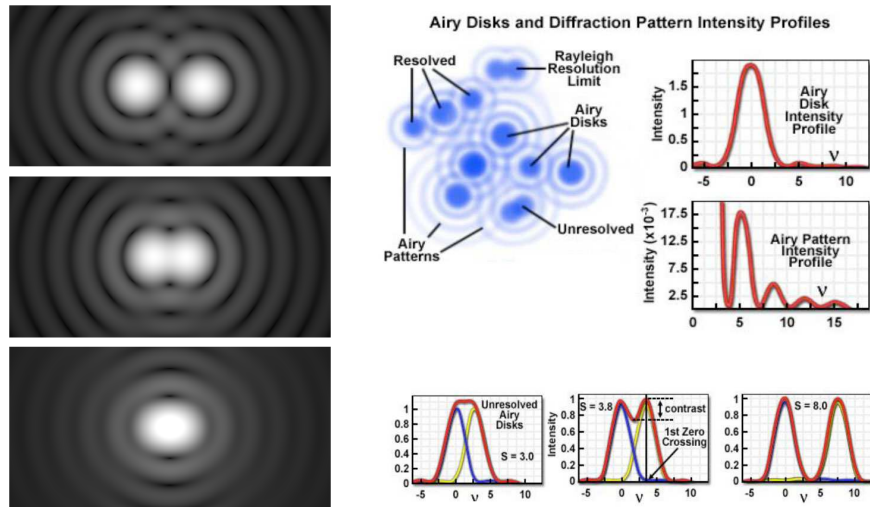
Numerische Apertur

Intensity Distribution of Diffracted Light

Figure 4

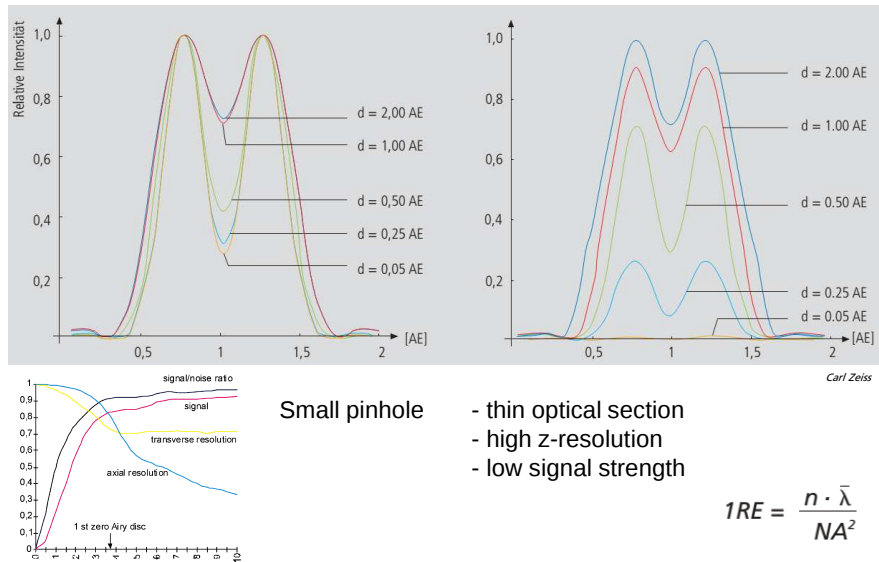


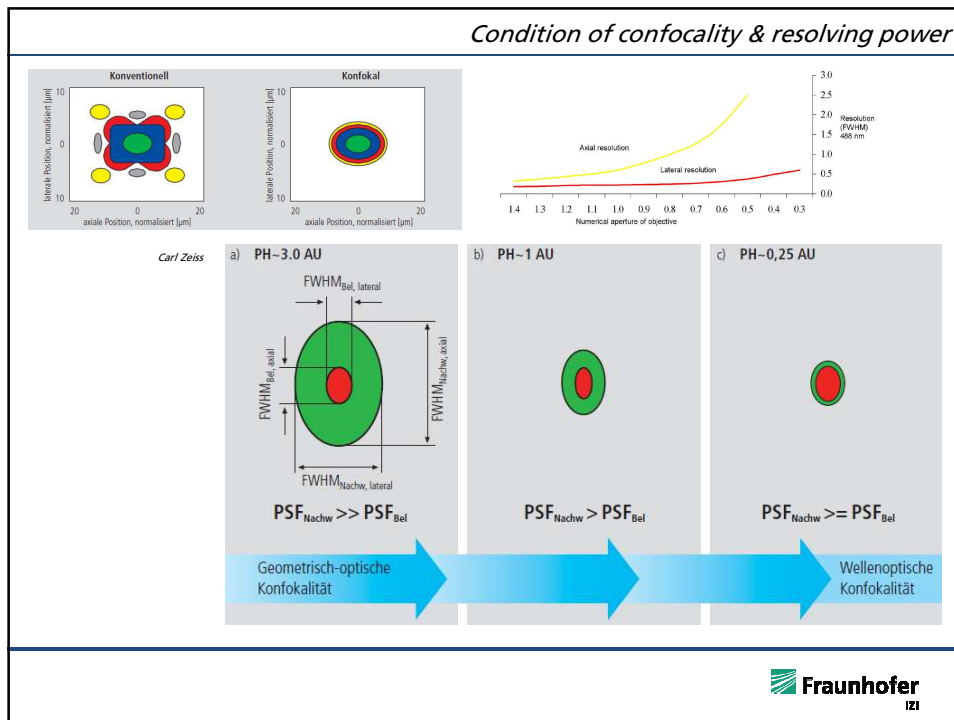
Angular resolution and Rayleigh criterion



26.5% depression in brightness btw two maxima is giving the sensation of twoness

Pinhole / Rayleigh unit and intensity





Resolution limits

Conventional Microscopy: $d = \frac{0,61 \cdot \lambda}{NA}$ d = optical resolution
 NA = Numerical aperture
 λ = Wavelength of light

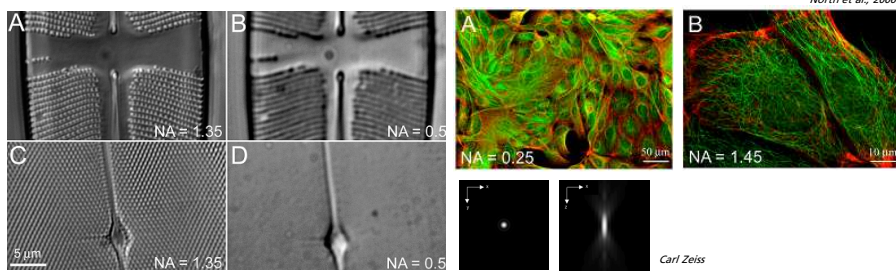
	Geometrisch optische Konfokalität	Wellenoptische Konfokalität	
Laterales Auflösungsvermögen	$\frac{0,51 \cdot \lambda_{em}}{NA}$	$\frac{0,37 \cdot \bar{\lambda}}{NA}$	Geometry Of the Probing beam spot
Axiales Auflösungsvermögen	$\frac{0,88 \cdot \lambda_{exc}}{n - \sqrt{n^2 - NA^2}}$	$\frac{0,64 \cdot \bar{\lambda}}{n - \sqrt{n^2 - NA^2}}$	Pinhole size
Optische Schnittdicke	$\sqrt{\left(\frac{0,88 \cdot \lambda_{em}}{n - \sqrt{n^2 - NA^2}}\right)^2 + \left(\frac{\sqrt{2} \cdot n \cdot PH}{NA}\right)^2}$	$\frac{0,64 \cdot \bar{\lambda}}{n - \sqrt{n^2 - NA^2}}$	

Fraunhofer
IZI

Numeric aperture and resolution

size varies: tissue / cellular / molecular - LSM limit

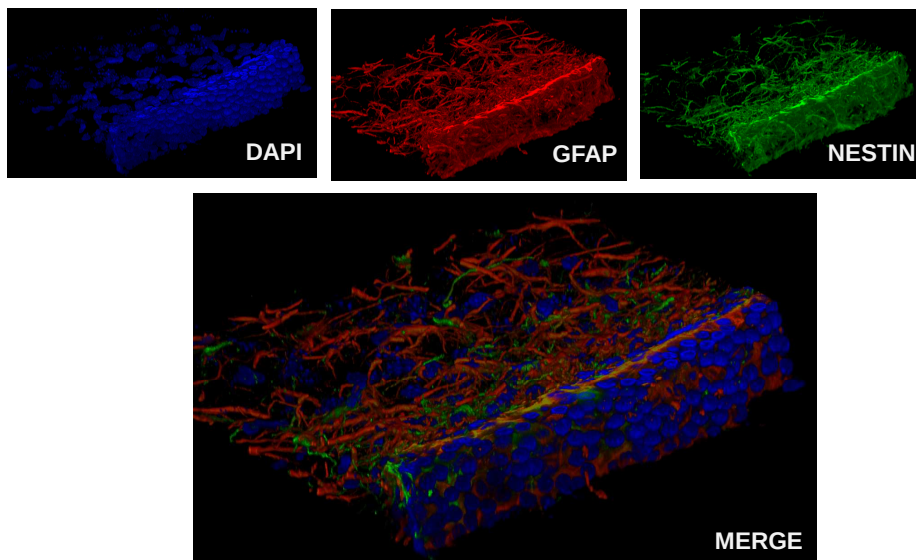
Animal cell: 10-30 μm	Nucleus: 3-10 μm	Microtubules: 25 nm
Antibody: 10-15 nm	GFP: 2-6 nm	Dye molecule: 1 nm



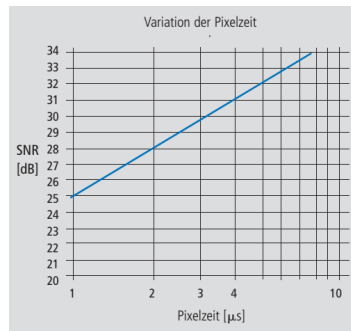
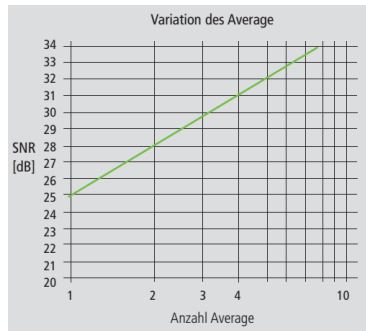
Light microscope best case
scenario: 200 x 200 x 400 nm

Check the PSF of the Lens!

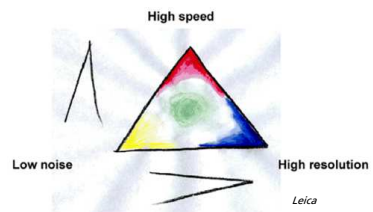
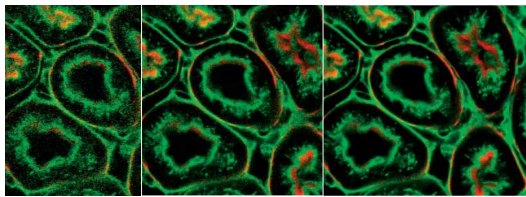
Example



Quality intensification

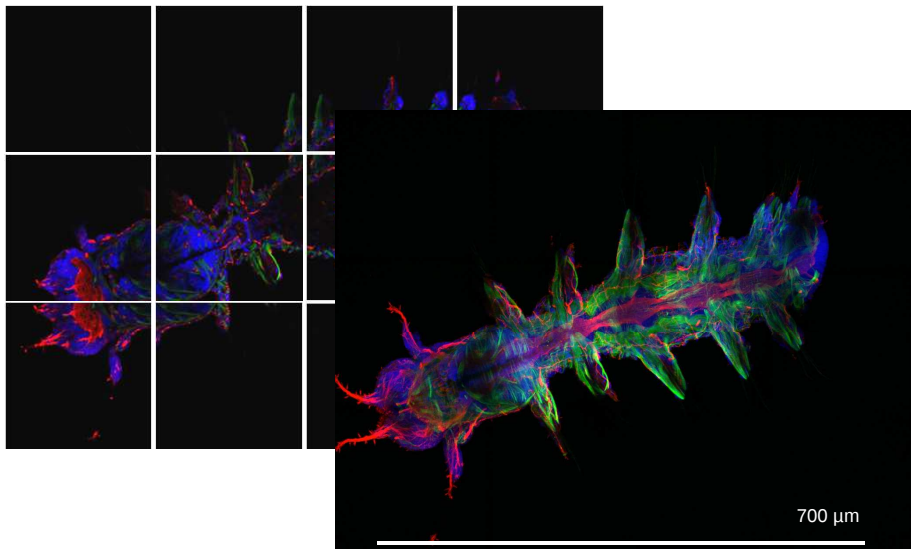


Carl Zeiss

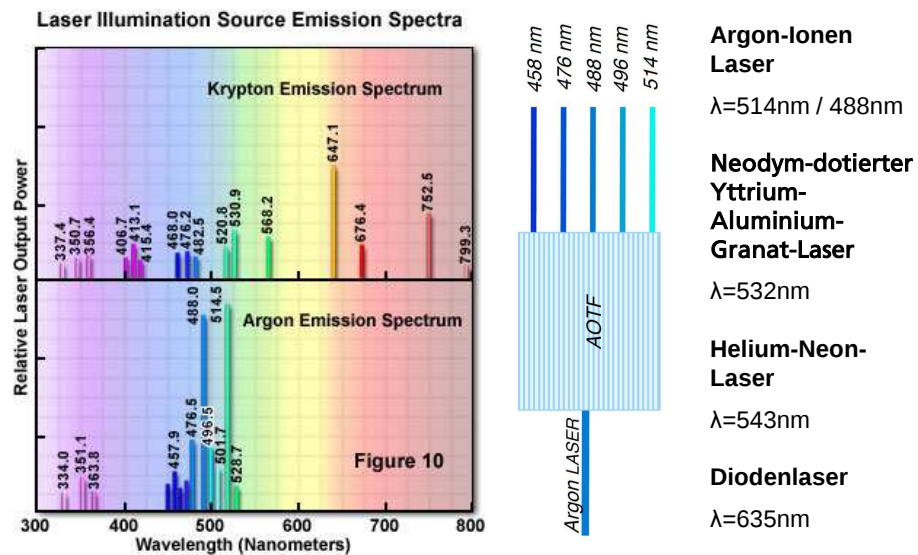
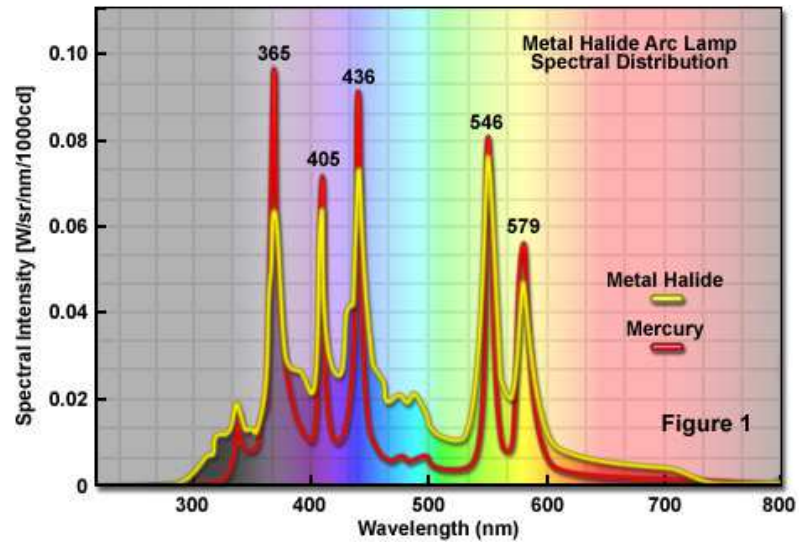


Fraunhofer
IZI

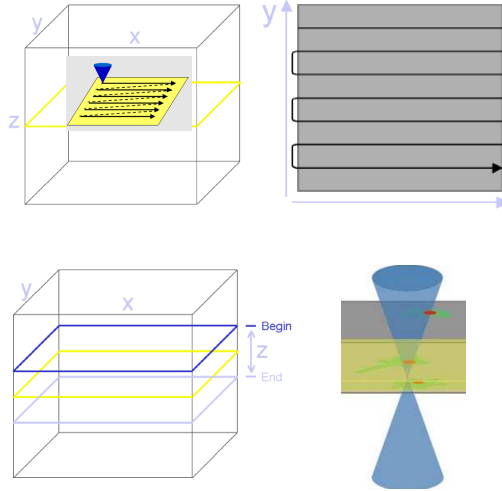
Stiching



Fraunhofer
IZI

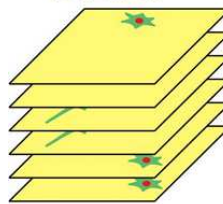


Beam scan:

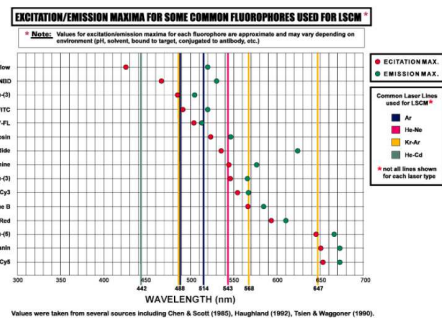
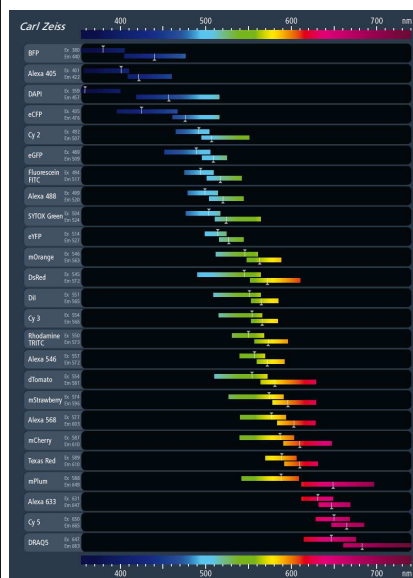


- x/y: by galvanometer driven scan mirrors
- Z: sample is moved via galvanometer driven stage or electronic focus of the microscope

X/Y/Z Stack



Fluorescent Dyes and Proteins



$$F = \sigma \cdot Q_e \cdot I \text{ [Photonen/sec]}$$

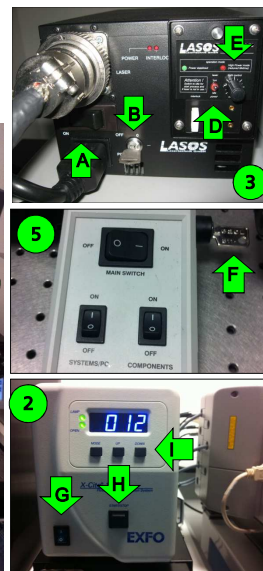
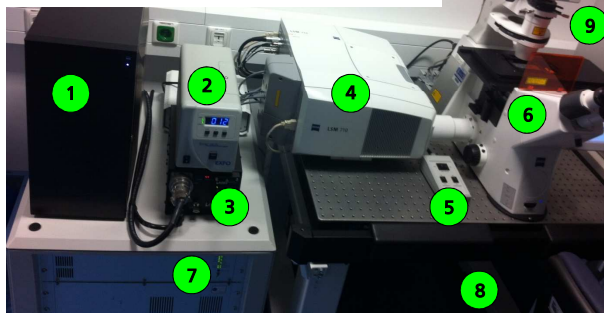
Our system



Fraunhofer
IZI

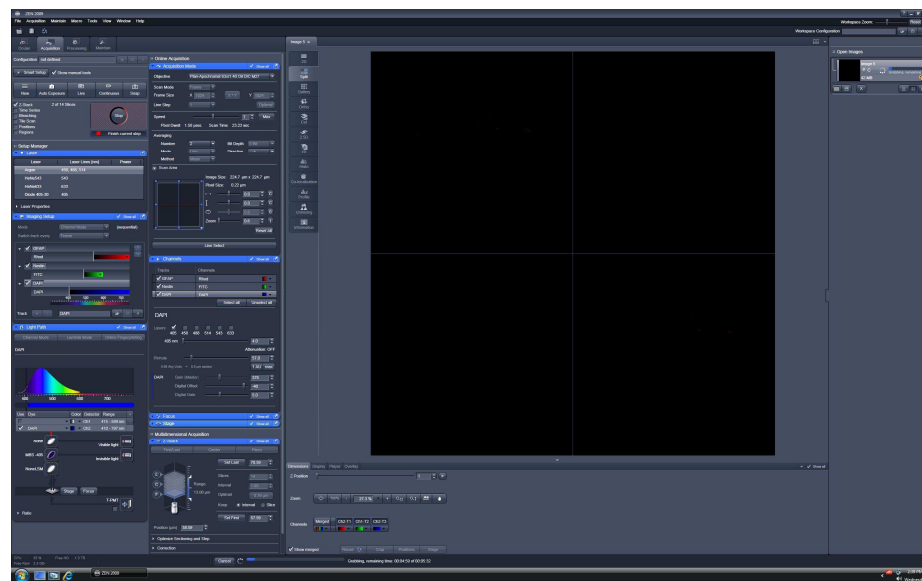
Our system

- | | | |
|------------------------------|-----------------------------|---|
| Server zur Datenverarbeitung | 1 Argon L. Hauptschalter | A |
| EXFO Fluoreszenzlampe | 2 Argon L. Power Key | B |
| Argon Laser | 3 Argon L. Power LED | C |
| LSM Modul am rechten Port | 4 Argon L. Idle/Run Switch | D |
| System Hauptschalter | 5 Argon L. Light Adjustment | E |
| Zeiss Axio Observer | 6 Hauptschalter Power Key | F |
| Laser Rack mit Leuchtdioden | 7 EXFO Hauptschalter | G |
| Steckerleiste unterm Tisch | 8 EXFO Shutter | H |
| Steuercomputer | 9 EXFO Setting Buttons | I |



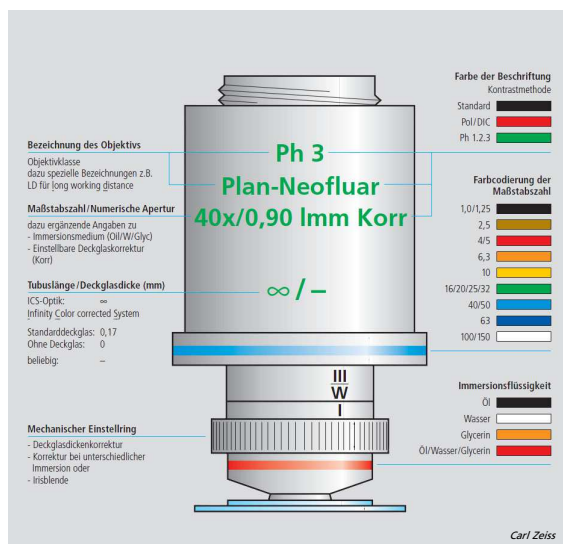
Fraunhofer
IZI

Our system: ZEN Software



Fraunhofer
IZI

Lenses



EC PLAN NEOFLUR

Korrektur 435nm – 670nm
3 Farb-Korrektur

10x 5.60mm 0.3NA T

20x T

40x 0.20mm 1.3NA ÖL DIC

PLAN APOCHROMAT

Korrektur 420nm – 670nm
4 Farb-Korrektur

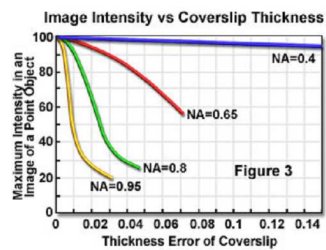
63x 0.18mm 1.4NA ÖL DIC

405 458 488 514 543 633

Fraunhofer
IZI

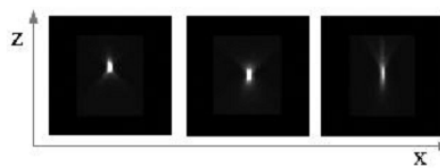
Correct coverslip thickness

The last 500 μm are important
Coverslip = 0.17 mm



Performance Reduction with Coverslip Thickness Variation

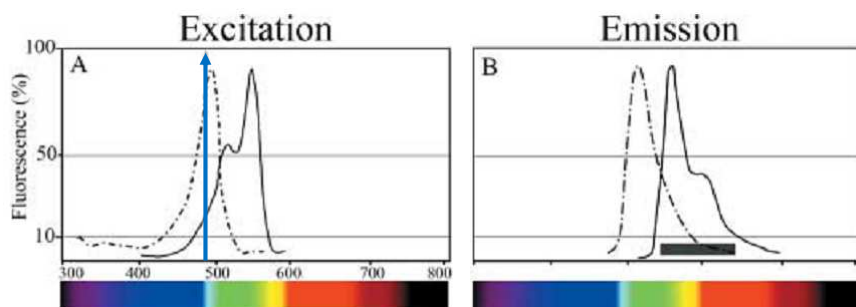
NumericalAperture	0.01 mm Deviation	0.02 mm Deviation
0.30	none	none
0.45	none	none
0.70	2 percent	8 percent
0.85	19 percent	57 percent
0.95	55 percent	71 percent



coverslip thickness: <0.17 mm 0.17 mm >0.17 mm

Diaz-Zamboni et al., 2008

Bleedthrough and crosstalk



Botte et al., 2006

FITC 490 nm max. Ex
Cy3 552 nm max. Ex
Laser 488nm

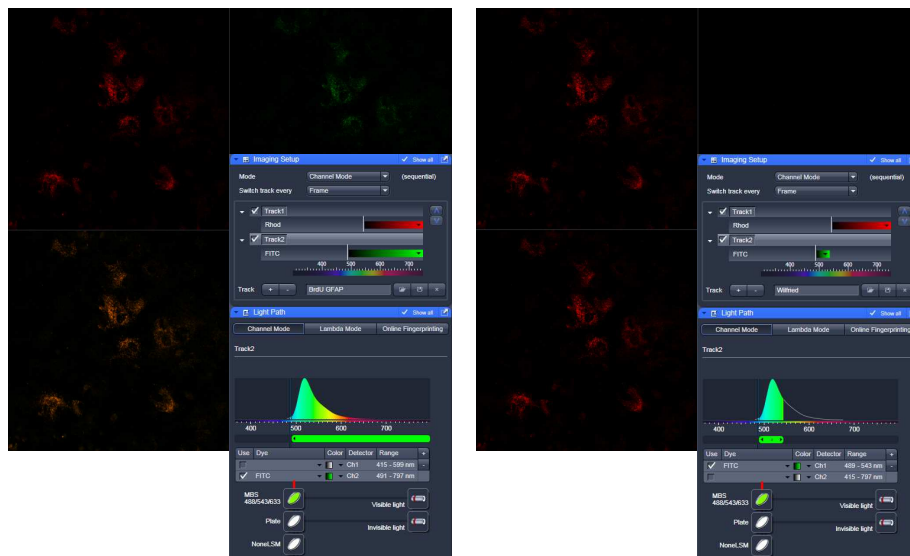
FITC 520 nm max. Emi
Cy3 570 nm max. Emi
PMT Cy3 Detection

Cross-talk

Bleed-through

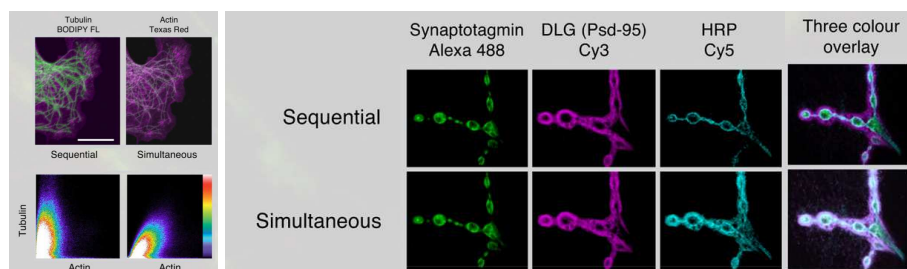
Always use sequential acquisition!

Bleedthrough

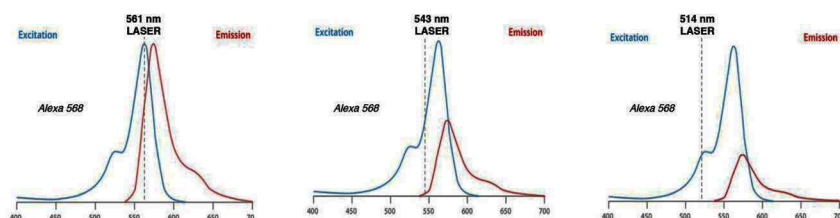


Fraunhofer
IZI

Cross-Talk & Link between Excitation / Emission



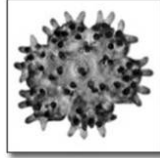
Brown et al., 2007



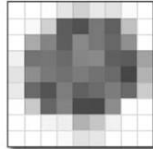
Remote excitation – Lower emission

Fraunhofer
IZI

Analog Image



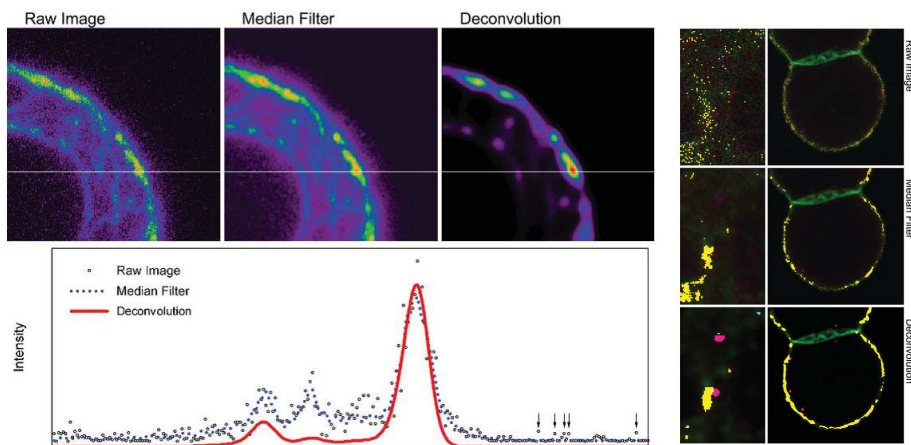
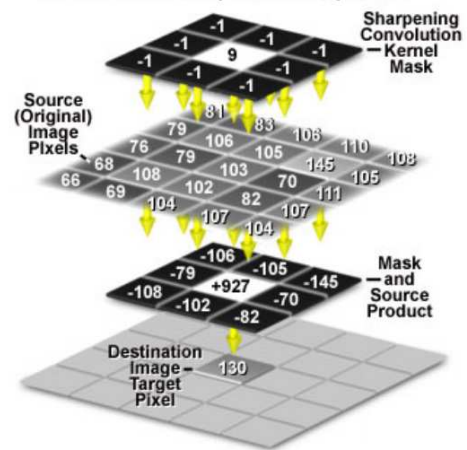
Digital Sampling



Pixel Quantization

249	244	240	230	209	233	227	281	288
248	249	210	85	81	120	87	193	254
280	170	133	94	137	120	104	148	283
241	116	118	107	134	138	96	82	183
277	142	121	113	124	118	107	71	179
234	106	84	126	87	108	128	108	204
241	202	102	132	78	73	141	248	282
283	282	244	239	179	199	242	280	246
288	249	244	280	229	231	240	281	283

The Convolution Operation Sequence



Landmann et al., 2004

Use Deconvolution

Calculation based on PSF of the lens

Digital File Format Memory Requirements

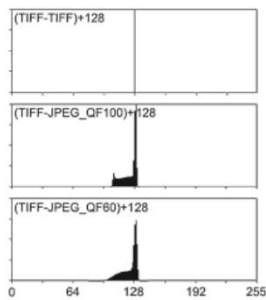
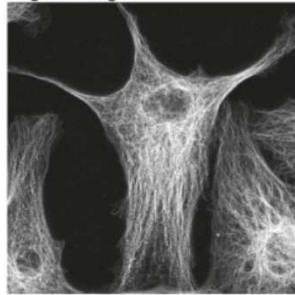
Pixel Dimensions	Grayscale (8-Bit)	Bitmap (24-Bit)	JPEG (24-Bit)	TIFF (24-Bit)
16 x 16	2k	2k	2k	2k
64 x 64	6k	13k	5k	13k
128 x 128	18k	49k	12k	49k
256 x 256	60k	193k	27k	193k
320 x 240	77k	226k	24k	226k
512 x 512	258k	769k	52k	770k
640 x 480	302k	901k	56k	902k
800 x 600	470k	1,407k	75k	1,408k
1024 x 768	770k	2,305k	104k	2,306k
1280 x 1024	1,282k	3,841k	147k	3,842k
1600 x 1200	1,877k	5,626k	161k	5,627k
2250 x 1800	3,960k	11,869k	276k	11,867k
3200 x 2560	8,002k	24,001k	458k	24,002k
3840 x 3072	11,522k	34,561k	611k	34,562k

JPEG = joint photographic experts group

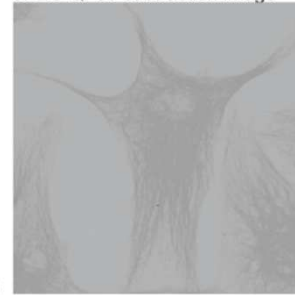
TIFF = tagged image file format

Use proprietary formats of your vendor

original image



JPEG QF60 subtraction image



Cromey et al., 2010

Bit Depth and Gray Levels in Digital Images

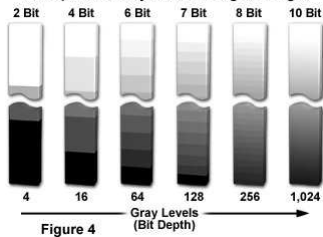
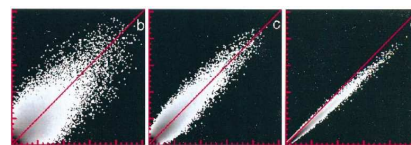
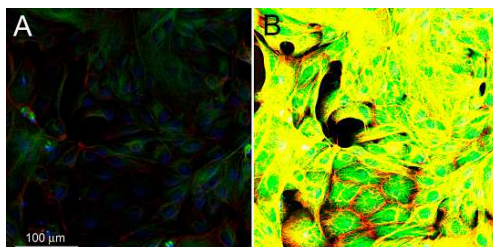


Figure 4



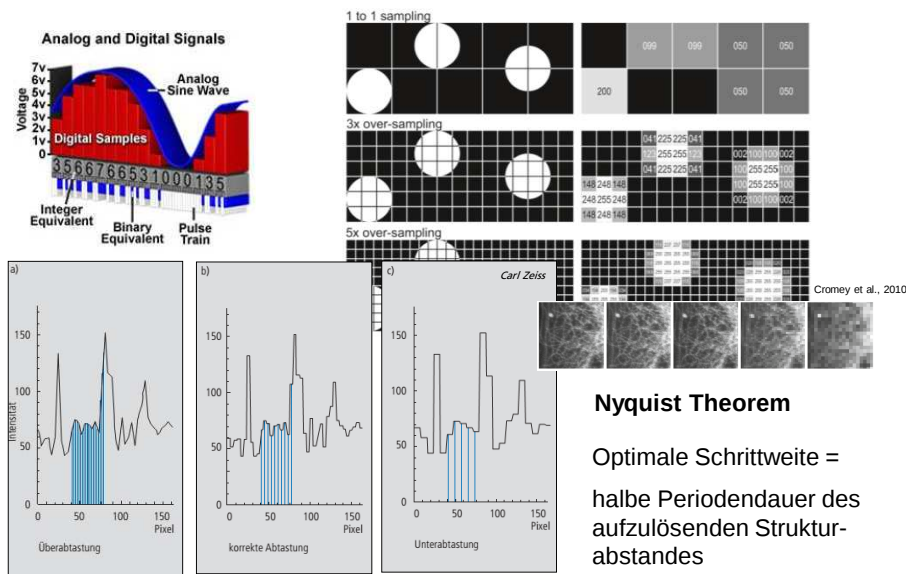
Demandolx et al., 1997

Dont bleach the area before imaging
-> bad S/N ratio



North et al., 2006

NA determines intensity
Avoid saturation! = data loss



Oversampling XY

Oversampling Z

Pinholes

1 Ary/ Channel

equal optical sections

Match all!

Table 1. The laws of Abbe and their effect on optical resolution and pixel sizes in wide-field and confocal microscopy.

	Wide-field		Confocal	
	Lateral resolution dx, y	Axial resolution dz, z	Lateral resolution dx, y	Axial resolution dz, z
Expression	$0.61 \lambda_w / NA$	$2 \lambda_w / NA^2$	$0.4 \lambda_w / NA$	$1.4 \lambda_w / NA^2$
Limit resolution of a 63x oil immersion objective with $NA = 1.32$ at $\lambda_w = 500$ nm	232 nm	574 nm	152 nm	402 nm
Minimal justified pixel size for this objective	101 nm	250 nm	66 nm	175 nm

Increasing the **brightness** shifts the entire intensity histogram to the right.
Increasing the **contrast** causes the intensity histogram to expand

Positive **gamma** (>1) increases the intensity of the mid-tones in the image more than the darker or lighter parts of the image.

Because gamma adjustments are non-linear, they should be declared in the figure legend or the methods section of a paper.

Brightness 😊

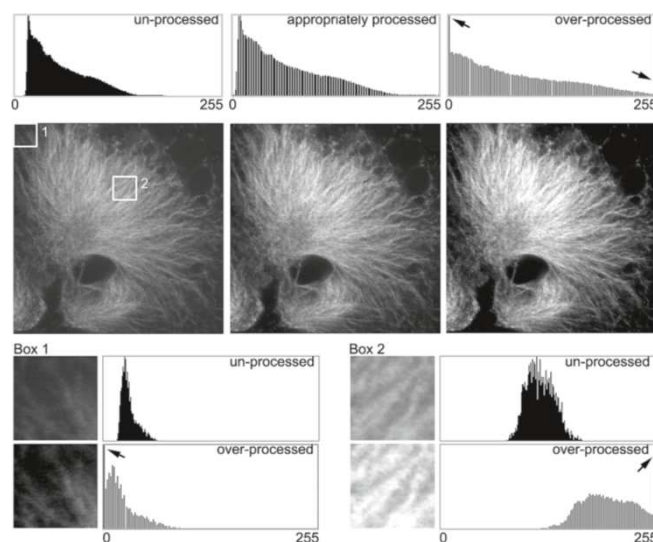
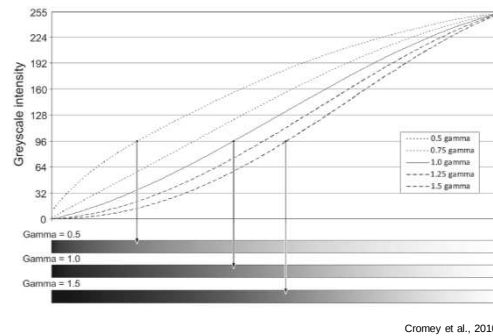
Contrast 😊

Gamma 😞

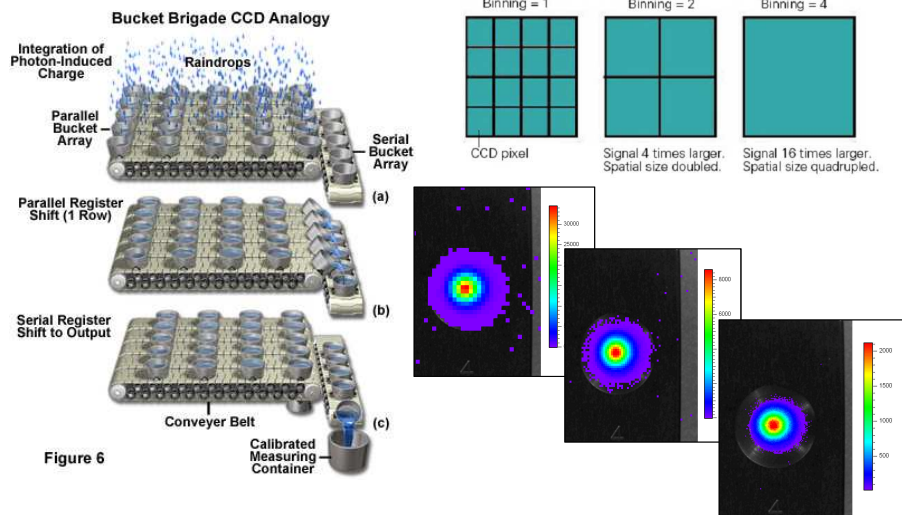
Carefull with filters!

Change whole picture

No selective changes

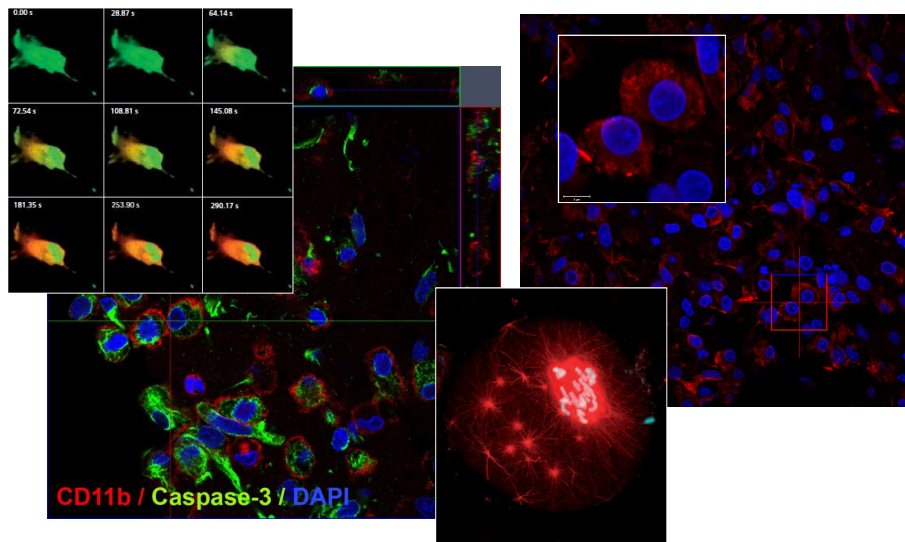


Pixel binning: basic principle

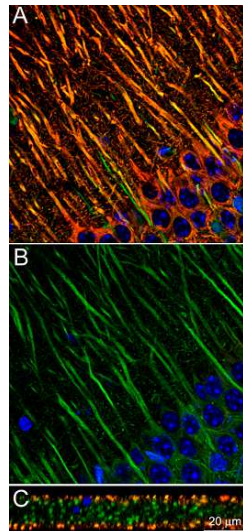


Fraunhofer
IZI

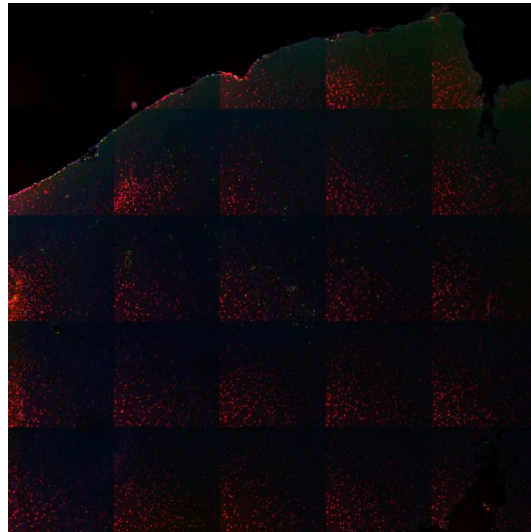
Examples



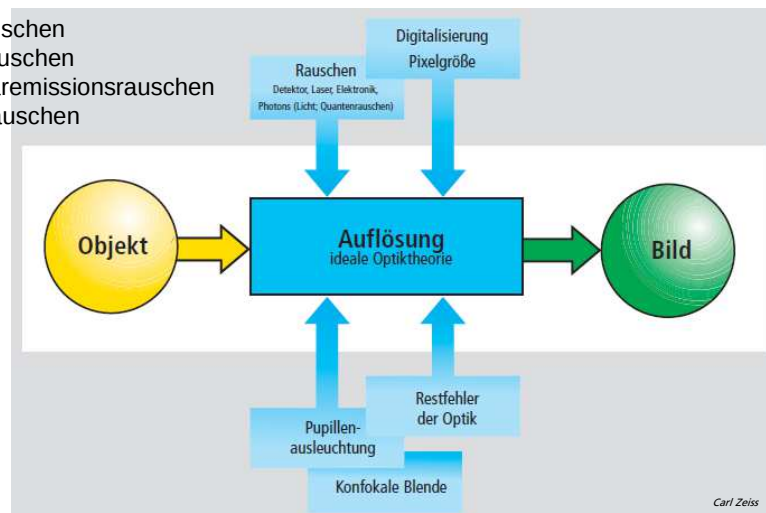
Fraunhofer
IZI



North et al., 2006



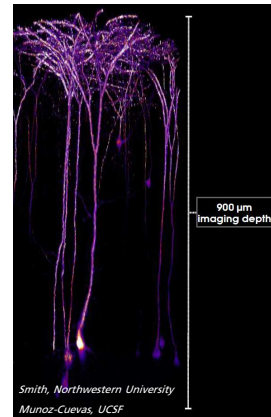
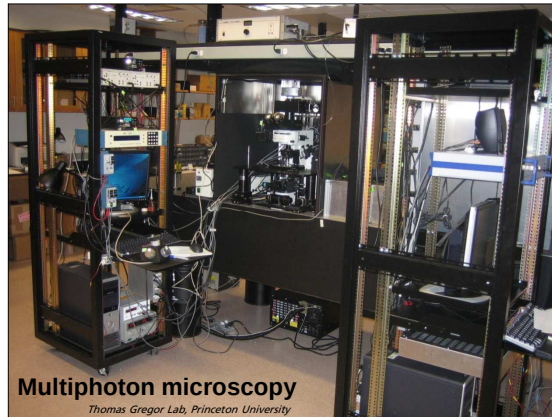
Laserrauschen
Schrotrauschen
Sekundäremissionsrauschen
Dunkelrauschen



Carl Zeiss

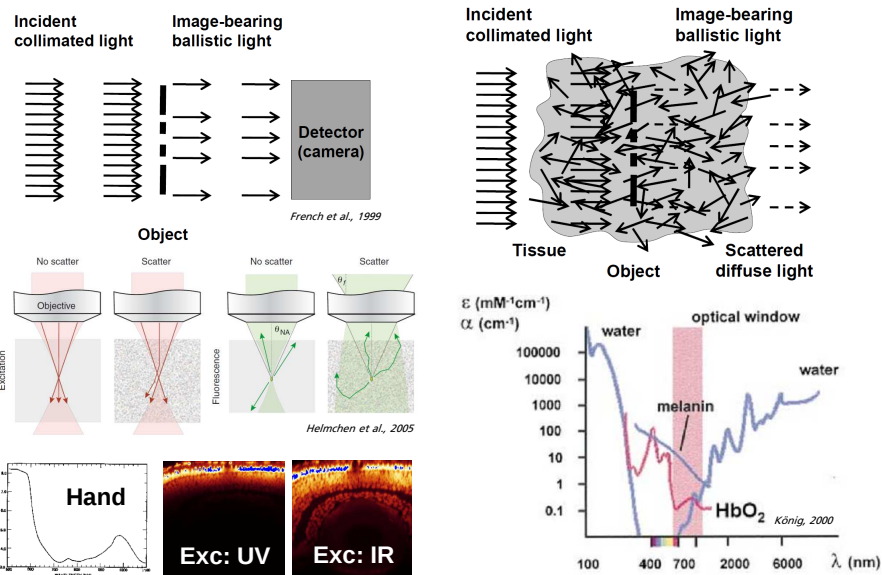
Fraunhofer IZI

Experimental Imaging - Alexander Kranz, MD

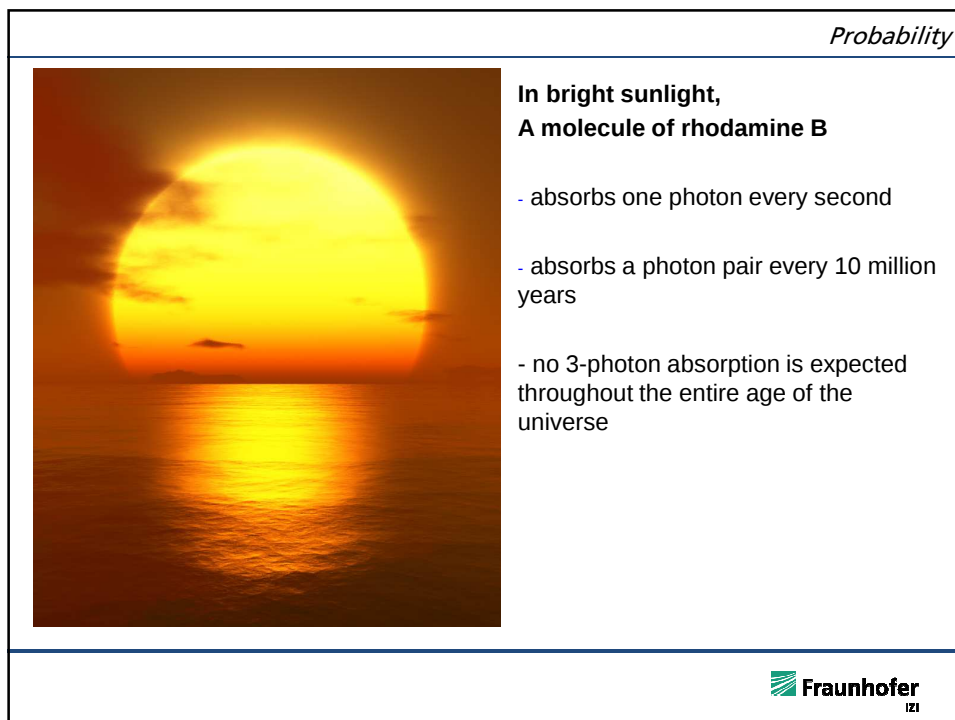
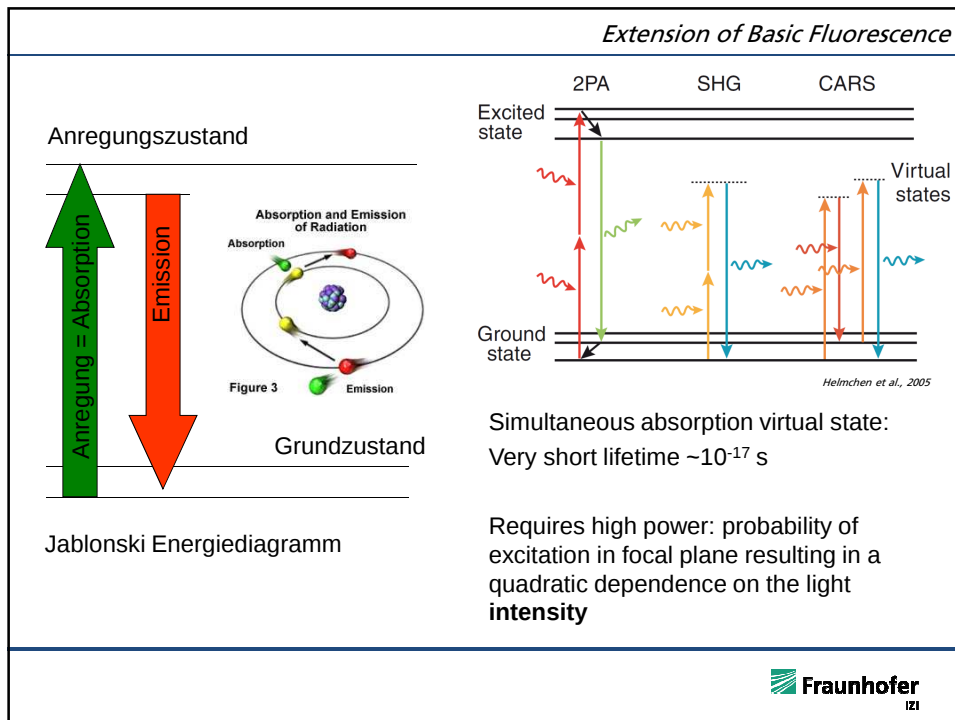


Fraunhofer
IZI

Light in tissue



Fraunhofer
IZI



History of Multiphoton Microscopy

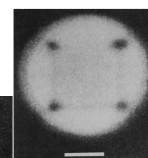
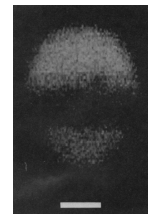
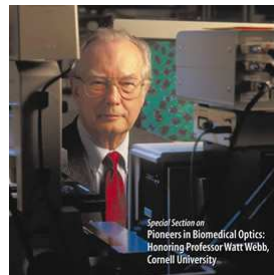


Göppert-Mayer
1931

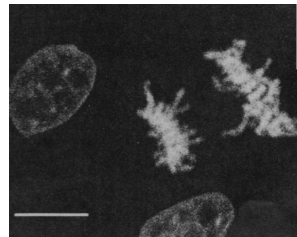
➔
Kaiser
1960



Denk & Webb
1990



Denk et al., 1990



Über Elementarakte mit zwei Quantensprüngen *Von Maria Göppert-Mayer*

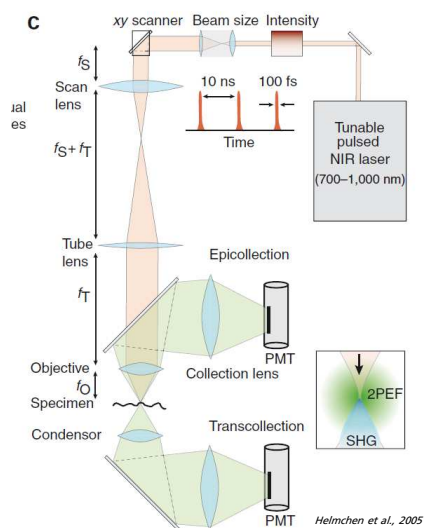
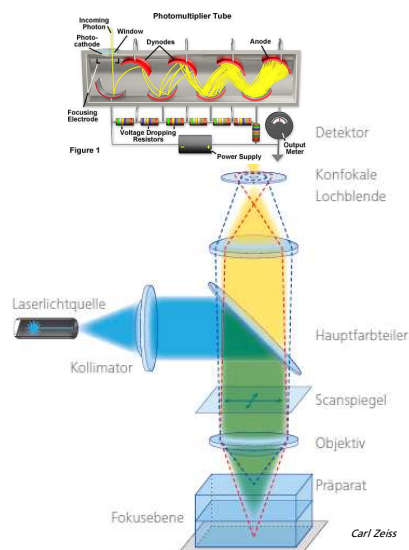
(Göttinger Dissertation)

(Mit 5 Figuren)

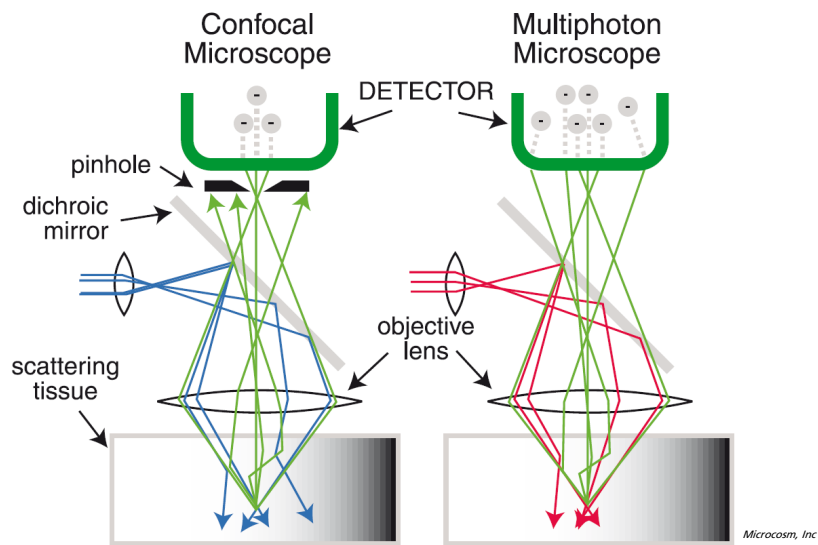
Einleitung

Der erste Teil dieser Arbeit beschäftigt sich mit dem Zusammenwirken zweier Lichtquanten in einem Elementarakt. Mit Hilfe der Diracschen Dispersionstheorie¹⁾ wird die Wahrscheinlichkeit eines dem Ramaneffekt analogen Prozesses, nämlich der Simultanemission zweier Lichtquanten, berechnet.

Comparison of structural composition



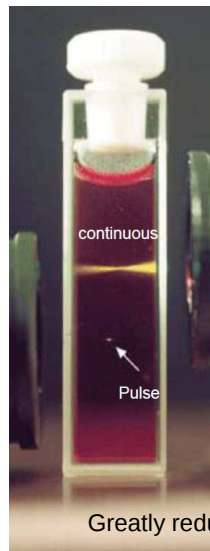
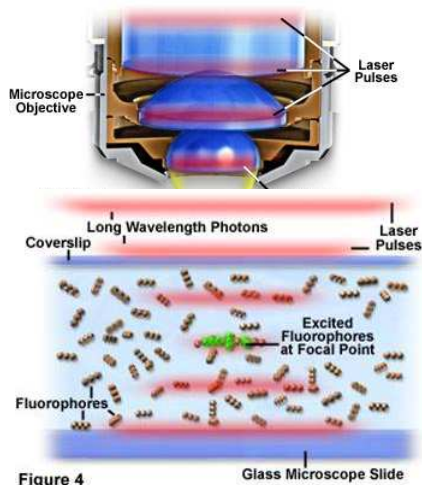
2PM: Intrinsic sectioning without pinhole aperture



Fraunhofer
IZI

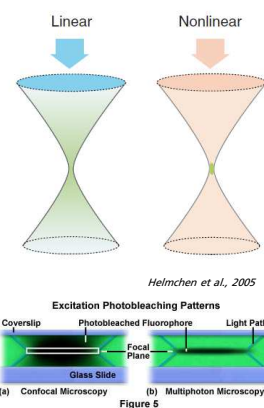
Focal spot multiphoton microscopy

Multiphoton Excitation Fluorescence Microscopy



Greatly reduces out of plain bleaching

Oheim et al., 2006



Fraunhofer
IZI

Overview Lasers

Laser Type (Spectral Region)

Wavelength(s) (Nanometers)

Argon Fluoride Excimer (UV)	193
Krypton Chloride Excimer (UV)	222
Helium Cadmium (UV, Visible)	325, 442
Argon (Visible)	488, 514
Copper Vapor (Visible)	510, 578
Nd:YAG Frequency Doubled (Visible)	532
Helium Neon (Visible, Near IR)	543, 594, 612, 633, 1150, 3390
Gold Vapor (Visible)	628
Rhodamine 6G Dye (Visible, Tunable)	570-650
Ruby (Visible)	694
Diode Semiconductor (Visible, Near IR)	630-1600
Ti:Sapphire (Visible - Near IR)	680-1130
Nd:YAG (Near IR)	1064
Erbium (Near IR)	1540
Hydrogen Fluoride (Near IR)	2600-3000
Carbon Dioxide (Far IR)	9600, 10600

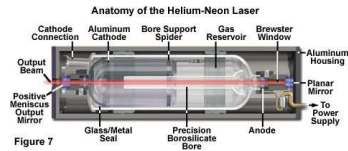


Figure 7

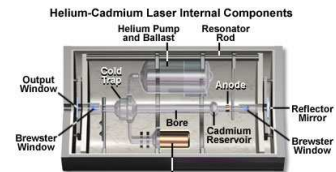
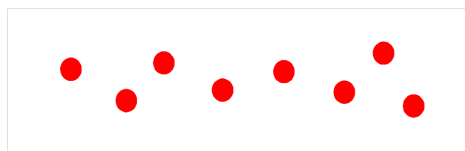


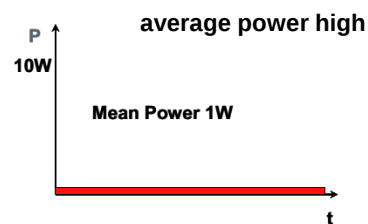
Figure 6

Laser Types

Continuous wave laser



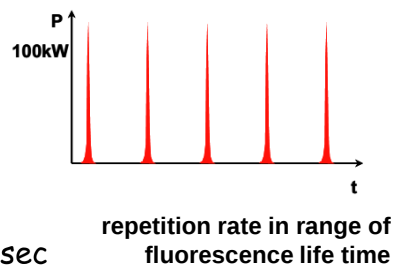
Time →



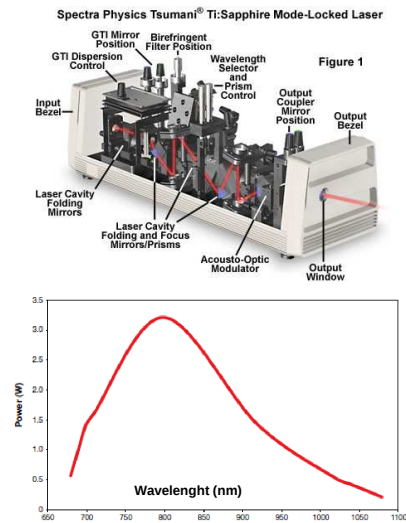
Pulsed laser



T 12.5nsec psec/fsec

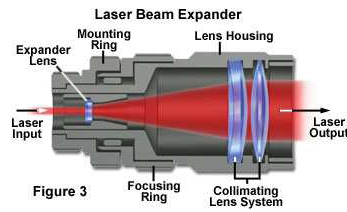
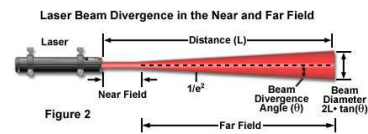


2PM: Cannot use continuous wave lasers



A laser for two photon microscopy:

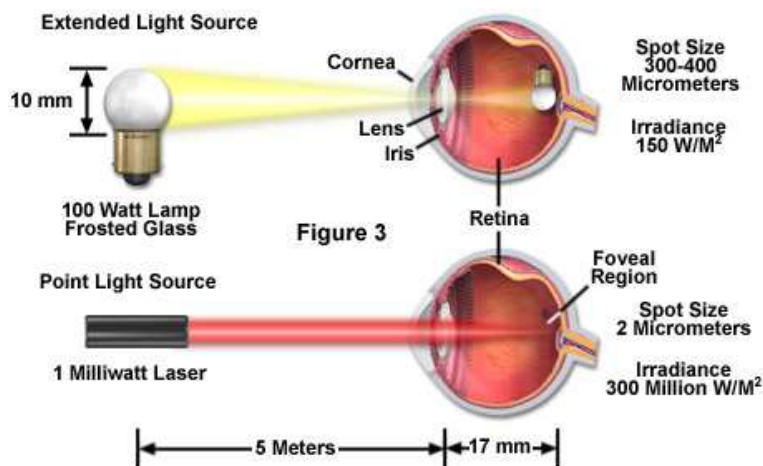
tuning range 690 to over 1050 nanometers
pulse widths ~ 100 femtoseconds
Pulse frequency 80 MHz
average power 2W



Most common MP laser is mode locked titanium sapphire (Ti:S), tunable 700-1000nm

Safety

Extended and Point Source Power Density at the Retina



Suitable dyes for Multiphoton

Fluorochrome	Absorption	Emission	Fluorochrome	Absorption	Emission
Alexa Fluor 350	720-800	440	DiA	800-860	580
Alexa Fluor 488	720-800	515	DID	780-820	670
Alexa Fluor 546	720-840	569	DIO	780-830	510
Alexa Fluor 568	720-840	596	eCFP	800-900	476
Alexa Fluor 594	720-850	610	eGFP	820-950	509
Alexa Fluor 633	720-900	647	eYFP	860-950	532
AMCA	780-800	444	Fluorescein	780 - 820	519
bis-MSB	680-750	420	Indo-1 free	690-720	490
Bodipy	900-950	512	Indo-1 Ca2+	690-720	400
Calcium Crimson	900	615	Lucifer Yellow	860-890	533
Calcium green	780-850	531	Mito Tracker red	750-840	600
Cascade Blue	750-800	420	Nile Red	810	640
Coumarin 307	780-800	530	Oregon Green	780-860	526
CY2	780-800	506	Propidium Iodide (PI)	820-850	617
CY3	780	565, 615	Rhodamin B	800-840	600
CY5	780-820	670	Rhodamine 123	780-860	550
Dansyl Hydrazine	700-750	440	Sytox Green	740-760 or 880-940	524
DAPI, Hoechst	700-820	455, 478	TRITC	800-840	572

Zeiss

Conventional dyes have a broad excitation spectrum when excited with pulsed NIR-L

Excitation probability

$$n_a \approx \delta \left(\frac{P_{avg}}{\tau f} \right)^2 \left(\pi \frac{NA^2}{hc\lambda} \right)^2$$

cross section = Wirkungsquerschnitt =
probability of a molecule to absorb
2 photons simultaneously

MP excitation favoured when:

- large cross section
- high laser peak power
- high NA objective lens
- low wavelength
- short puls width and repetition time

n_a : probability of excitation

δ : excitation cross section

**P_{avg} : average power incident light
(peak power)**

τ : pulsewidth

f : repetition rate

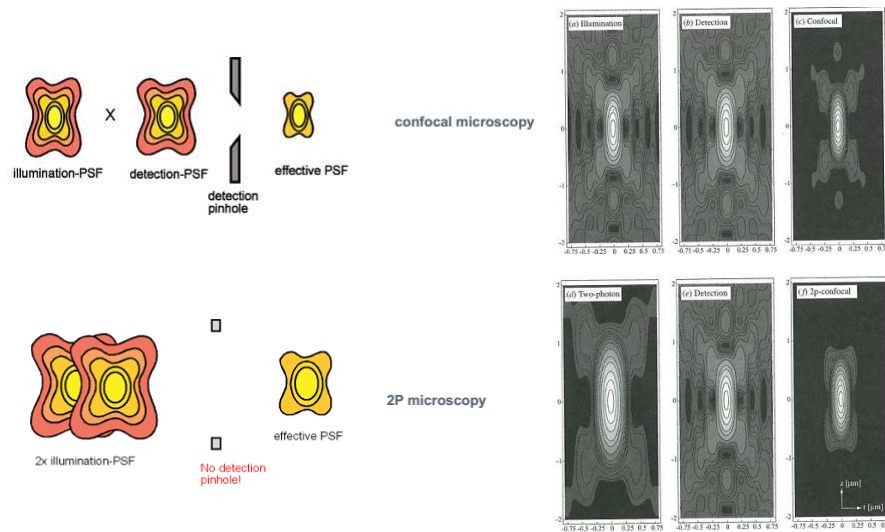
NA : Numerical aperture

h : Planck's constant

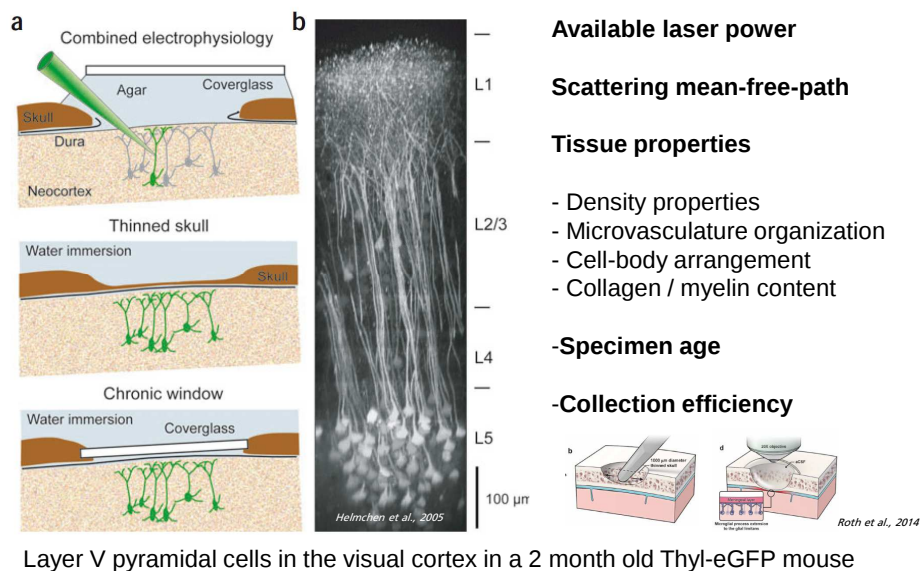
c : Speed of light

λ : Wavelength

Comparing confocal and multiphoton



Maximum imaging depth



Summary Multi Photon Microscopy

True „Molecular Imaging“ with single-molecule sensitivity

Wealth of indicators capable of specific targeting (dyes, SHG, quantum dots)

Sub-micron resolution

Optical sectioning in thick, turbid media and deep penetration

Wide variety of biological and clinical applications

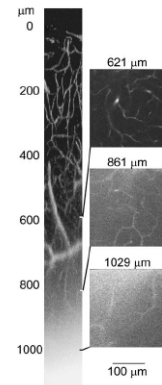
Near IR-light scatters less and is less toxic than blue light

Limited photobleaching and photodamage in the image plane

More efficient light collection (better looking images)

Unaffected by chromatic aberrations

High purchasing and operating cost of pulsed femtosecond IR lasers



Svoboda et al., 2006

Source

<http://www.microscopyu.com/>

<http://zeiss-campus.magnet.fsu.edu/>

<http://www.olympusmicro.com/>

<http://www.leica-microsystems.com/science-lab/>